

<b>Title</b>	DNA extraction from rapid diagnostic tests (RDT) by Chelex Method
<b>SOP code</b>	SOP_LAB_Pf_62_v01
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## 1 Objectives

Describe the procedures for the extraction of DNA from blood onto nitrocellulose strips in rapid diagnostic tests (RDT) using the Chelex-based method.

## 2 Definitions

RDT: Rapid Diagnostic Test

PBS: Phosphate-buffered saline

## 3 Responsibilities

This procedure is designed for use by appropriately-equipped laboratories. Training is required to perform the procedure successfully. Trained researchers/laboratory technician are responsible of the correct execution of the SOP. The principal investigator and co-principal investigator are responsible for approval and evaluation of the procedures

## 4 Related SOPs

- SOP\_LAB\_BCN\_59\_v01\_EN (2017): DNA extraction from dried blood spots by Chelex method
- SOP\_LAB\_BCN\_64\_v01\_EN (2023) Extraction of DNA from dried blood spots (DBS) and rapid diagnostic tests (RDT) by Chelex method in 96 well plates
- SOP\_LAB\_BCN\_19\_v02\_EN (2018): Detection and quantification of *Plasmodium falciparum* by Real Time PCR

## 5 Procedures

### 5.1 Materials required

#### a. Reagents

- PBS.
- Chelex® 100 sodium form (C7901, Merck).
- Nuclease free water
- Bleach
- Ethanol

#### b. Consumables

- Filter tips (10 µL, 200 µL and 1000 µL)
- Wide opening pipette tips: 200µl (Ref: 541152-965-008-09, Cultek).
- 1.5 mL centrifuge tubes
- Disposable gloves
- Fine tip marker pens

- 50ml tubes
- Plastic sterile Pasteur pipettes

#### c. Reference Samples

- Plasmodium infected whole blood in RDT nitrocellulose strips (RDT).
- Uninfected blood on RDT as negative controls

#### d. Equipment

- Vortex, mini-centrifuge
- P1000, P200, P20, P10 pipettman
- Dry bath
- Scissors

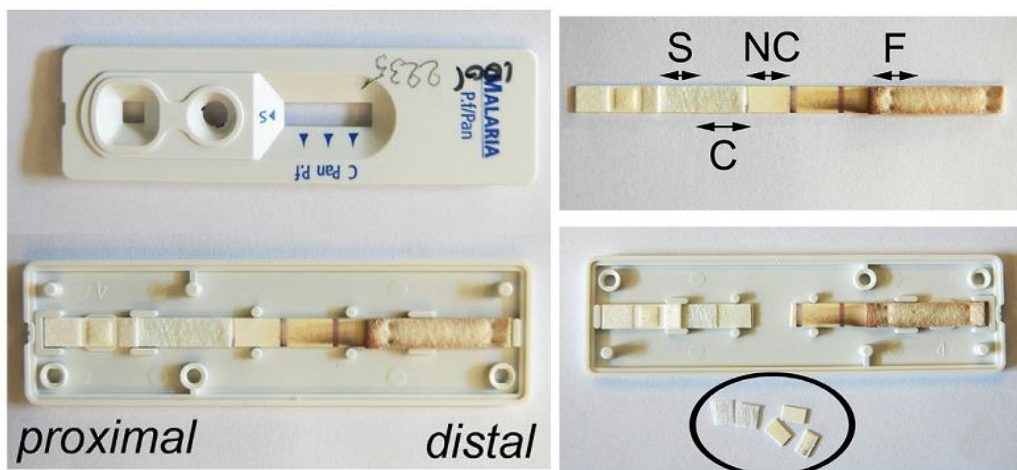
### 5.2 Preparation of solutions and buffers (before starting the procedure)

- 20% Chelex: weigh 2g of Chelex 100 sodium form and mix into 10ml of H<sub>2</sub>O<sub>dd</sub>. Store at 4°C.
- 5% bleach in distilled water
- 70% ethanol in distilled water
- Filtered PBS.

### 5.3 Working procedure

- Clean the scissors and forceps after processing each RDT by dipping 1min in 5% bleach and 1min in 70% ethanol using 50ml falcon tubes or similar containers, and let them dry.
- Open the RDT cassette using sterile scissors and forceps, and remove the nitrocellulose strip.
- Cut the conjugate pad (C) together with the proximal part of the nitrocellulose strip (NC) (see Figure 1)

Figure 1:



- Put the pieces of the nitrocellulose strip in the bottom of a 1,5ml eppendorf.
- If extracting DNA from multiple RDT cassettes, clean the scissors and forceps after processing each RDT by dipping 1min to 5% bleach and 1min to 70% ethanol.
- Add 1 ml PBS to the 1,5ml eppendorf with the pieces of nitrocellulose. Ensure the sample are soaked in buffer. Vortex 5s.
- Incubate at room temperature for 10 min.
- Centrifuge at 14,000 rpm for 2 min and discard supernatant using a clean pipette tip for each sample.

- i. Add 1 ml PBS. *Vortex 5s.*
- j. Centrifuge at 14,000 rpm for 2 min and discard supernatant using a clean pipette tip for each sample.
- k. Create a 1:2 solution of Chelex in nuclease-free water (for example, take 1mL 20% Chelex + 2mL H<sub>2</sub>O)
- l. Add 100µl of 1:2 Chelex solution to the eppendorf using a wide opening pipette tip (200µl)  
*Note: Mix very well the Chelex solutions before use it and do it quickly- Chelex solution is very heterogeneous and Chelex beads tend to settle on the bottom very quickly.*
- m. Incubate at 95 °C for 10 min.
- n. Centrifuge at 14,000 rpm for 1 min.
- o. Transfer supernatant into a clean microcentrifuge tube.  
*Note: Try not to transfer Chelex beads as it may inhibit the qPCR.*
- p. Store supernatants at +4°C and do the qPCR. If storing samples for longer than a day, store at -20 °C.

## 6 References

- Cnops, L., Boderie, M., Gillet, P., Van Esbroeck, M., and Jacobs, J. (2011). Rapid diagnostic tests as a source of DNA for Plasmodium species-specific real-time PCR. *Malaria Journal* 10.
- WWARN (2014). Preparation of Rapid Diagnostic Tests (RDTs) for DNA extraction. 1.1 1–7.